Fewer insect and mite pests attack grapes grown in British Columbia compared with many other major grape producing regions, which allows for a ‘softer’ approach to pest management that preserves beneficial insects and predacious mites that help regulate secondary pests such as spider mites and grape mealybug. Adoption of integrated pest management (IPM) practices that minimize the use of chemical sprays can also help reduce production costs, reduce human exposure to insecticides, and preserve the local environment. Insect and Mite Pests of Grape in British Columbia (Lowery et al., 2014), a companion publication contains colour photographs of many of the pests outlined in this chapter of the production guide. In the following descriptions of grapevine pests, numbers and letters indicated in parentheses refer to the pages and plates of the corresponding photo guide. Additional sources for information on insect and mite pests of grapes are listed at the end of this section.

**Integrated Pest Management**

IPM utilizes a number of principles and practices to manage pest populations, including biological, cultural, and physical control methods. Insecticides remain an essential component of an IPM program, but they are applied as a last resort only when monitoring has shown that pest numbers are likely to exceed the economic threshold. When insecticides are required, their selectivity, persistence, and effects on non-target organisms should be considered. Some have minimal effects on beneficial insects and predacious mites, while others cause significant reductions in natural enemy populations. A brief description and a few examples of the various components of a grape IPM program are outlined below. For additional information, refer to the Integrated Pest Management section (5.1) of this guide.

**Biological Control of Insects and Mites**

Insect and mite pests of grapes are attacked by many species of beneficial organisms, including bacterial and viral diseases, spiders, insect predators, predacious mites, and vertebrates such as toads and birds. Maintaining and enhancing numbers of these natural enemies of grape pests forms the cornerstone of a successful IPM program. Healthy populations of predators and parasites prevent outbreaks of secondary pests and reduce the numbers of sprays required for the control of primary pests such as leafhoppers and climbing cutworm.

Beneficial organisms can be preserved or enhanced in several ways. The negative impact of pesticides can be minimized by spraying only when and where required. Monitoring of pest numbers will often indicate that only a small portion of a vineyard requires chemical treatment. Selecting the most appropriate spray material will help reduce damage to non-target organisms. Compared with broad-spectrum insecticides that are often more toxic to beneficial insects than the pests they are intended to control, insecticides that are less damaging to beneficial insects and predacious mites (e.g. Altacor™) should be chosen whenever possible. Malathion is considered to be less damaging to beneficial insects and mites than other insecticides in the same class (organophosphates), and carbaryl (Sevin XL™) can be used at a low rate when the target is the susceptible Virginia creeper leafhopper rather than the more resistant western grape leafhopper. Other materials, such as the microbial insecticide Dipel (B.t.), are highly selective and require that the pest consumes the treated plant part, reducing toxicity to most non-target species. As listed in the BCMAL fact sheet ‘Pest Control Products Recommended for Use on Grapes in British Columbia’ (http://www.agf.gov.bc.ca/cropprot/grapeipm/grape_pesticides.pdf), all of the Class 4 (nicotinoid) insecticides (Assail™, Clutch™, Closer™) combined should not be applied more than twice per season to avoid mite outbreaks.
These materials are listed as toxic to bees. The Class 3 pyrethroids (Pounce™, Ambush™, Ripcord™) and natural pyrethrins (Pyganic™) are toxic to most beneficial insects, but they persist in the environment for only a short time.

Thoughtful choice of management practices can also help preserve beneficial insects. Mowing less often and mowing only alternate rows at one time are simple and cost effective ways to increase populations of most natural enemies. Adults of many beneficial species feed on nectar of flowering plants that also serve as hosts for alternate prey species. As a general rule, increased plant diversity is associated with greater numbers of beneficial insects. A diversity of plants can be provided in the vineyard in mixed ground covers, or in hedgerows and uncultivated areas within or adjacent to the vineyard.

Cultural and Physical Controls

Vine Vigour and Resistance

For most crops, varieties can be selected that are partially or wholly resistant to one or more pests and diseases. Except for rootstocks that are resistant to grape phylloxera or nematodes, this is generally not a viable option for wine grapes. Most desirable wine grape varieties possess little resistance to foliar feeding pests. Vine vigour does, however, influence insect and mite numbers. Leafhoppers and grape mealybug, for example, will reach significantly higher numbers on overly vigorous vines as they prefer the darker, sheltered environment and elevated humidity that excessive vine growth provides. Insects developing on these plants survive better and grow faster due to better nutrition, softer tissues, and changes in concentrations of secondary plant compounds. At the other extreme, chlorotic vines with low vigour are less able to tolerate insect feeding damage and are more susceptible to attack by hard scale insects and wood boring beetles. Fortunately, the optimum balance in vine growth that results in the highest quality wines is also best for minimizing the growth of pest populations.

Management of vine vigour is an important consideration in the establishment of a new vineyard and is discussed elsewhere. After vines are established, growth is controlled by pruning, cropping, selection and management of appropriate ground-cover plants, and the provision of water and nutrients. Recent research in California has shown that deficit irrigation from berry set to veraison reduced leafhopper numbers by more than 60%. The modest reduction in yield, around 15%, was more than offset by a significant improvement in wine quality. In addition, berries were smaller and exposed to more light and air and pruning costs were slightly lower.

The presence of broadleaf weeds in and between vine rows is associated with lower levels of cutworm damage. In spring, when possible avoid controlling shepherd’s purse and other weeds in the vine row until after buds have broken, as removal of these alternate sources of food forces cutworm larvae to feed more on the buds of grapes.

Pruning

Pruning can be altered to help reduce damage from cutworm larvae. Slightly more buds can be left on vines to compensate for damage, but this will require the removal of more unwanted shoots later in the season. Some growers delay suckering and shoot thinning to divert some leafhopper feeding and egg-laying to these unwanted plant parts. Thinning of the canopy by shoot removal, shoot positioning, and removal of basal leaves improves air flow and light penetration, which is important for the management of diseases as well as insects.

Leafhoppers and erineum or blister mite infest the first leaves that emerge in spring, and these lower leaves can be removed in June to reduce numbers of these pests. A study conducted in commercial British Columbia vineyards showed that removal of basal leaves in June rather than August reduced numbers of leafhoppers and the incidence of bunch rot. Vine vigour and berry size were reduced only slightly and there was little effect on ripening or quality of fruit. Early season leaf removal might not be suitable for stressed vines or vines on sandy sites with intense heat and light where fruit can become sunburned. As for late season removal, partial removal of leaves from only the shaded sides of the vines might be more suitable in these areas.

Physical controls

For some vineyards, the use of yellow sticky tape applied below the cordon in spring can be an economical way to manage leafhoppers.
Although costly, this physical control method preserves beneficial insects and reduces or eliminates the need for additional insecticides later in the season. Use of yellow sticky tape is most practical in vineyards where damaging numbers of leafhoppers occur in small, isolated areas or on a few outer rows of the vineyard. Other physical controls include pruning out of canes infested with scale insects and application of barriers to prevent climbing cutworm from reaching developing buds.

**Monitoring and Chemical Controls**

Insecticides should only be applied when monitoring indicates that sprays are warranted. The mere presence of a pest does not indicate that a spray is required. Established grapevines can tolerate moderate amounts of leaf damage and small numbers of pests may have little or no effect on vine growth and yields. Even when pests have reached damaging levels sprays should be withheld until the economic benefit of treating the crop exceeds the cost of controlling the pest, including possible disruption of beneficial insects. In other words, sprays should only be applied when pest populations reach the economic injury level where control of the pest provides an economic return to the grower.

Thresholds for grape pests are currently based mostly on visual inspections. Yellow sticky cards can be useful indicators of general population trends and pest developmental stages. Care should be taken to conduct a thorough inspection that takes into account the variable nature of the crop and the uneven distribution of most grape pests. Particular attention should be given to field edges and to areas that have experienced damage previously. The use of double sided clear tape applied around shoots or the vine cordon can be used to assist with timing of sprays for the control of the young motile stages of grape mealybug and soft scale insects.

Grapevines need to be monitored for pests throughout most of the year. It is easier to spot scale infestations during pruning or early in spring before leaves are present. Grapes need to be visually inspected frequently in spring when unopened buds are susceptible to cutworm damage. When the first leaves have fully expanded, watch for early infestations of erineum mite and note the presence of leafhopper adults that begin laying eggs at this time. Thrips damage table grapes from 75% bloom to fruit set, and if scale or mealybug were noted earlier, sprays can be timed based on the presence of crawlers. The need to spray for first generation leafhoppers should be based on monitoring for small nymphs from mid June to mid July, while second generation nymphs will reach their peak usually after the first week of August. In summer, the presence of soft scale and mealybug is often betrayed by the presence of honeydew and attendant ants. Throughout summer, monitor for thrips, spider mites and grape leaf rust mite. For table grapes, watch for mealybug infesting clusters in mid-summer and for smallcase bagworm and earwigs closer to harvest. Begin monitoring for spotted wing drosophila when fruit begins to ripen.

If chemical control of a pest is indicated, refer to the fact sheet Pest Control Products Recommended for Use on Grapes in British Columbia, posted at http://www.agf.gov.bc.ca/cropprot/grapeipm/grape_pesticides.pdf for suitable spray materials, or consult with private consultants or your chemical supplier. It is the legal responsibility of pesticide applicators to follow the label instructions and apply only those products that are registered for use on grapes. The British Columbia Wine Grape Council and the authors of these pest management sections do not assume legal responsibility for the misuse of any pesticides mentioned in this guide. For grapes or wines destined for export, check to ensure that the importing country has maximum residue levels (MRL) for the control products. Observe the days to harvest interval (PHI = pre-harvest interval) to avoid exceeding the MRL. Producers of organic grapes should also refer to the list of acceptable products and consult with an organic growers’ organization.

**Major Pests of Grape**

The major insect and mite pests of grapes grown in BC are cutworm larvae that attack buds in spring, leafhoppers that feed on leaves throughout the summer, wasps that eat ripe fruit and annoy or sting workers, and mealybug and soft scale that transmit grapevine viruses. Grape phylloxera is an important pest of grapes.
worldwide but it occurs in only a few scattered locations in the southern interior. Other than wasps, these pests are largely absent from the Lower Mainland and Vancouver Island. In addition to the major pests listed above, producers of table grapes need to worry about spotted wing drosophila, earwigs and scarring of fruit by thrips. Earwigs do not cause direct damage to grapes but are considered a contaminant by many vendors of table grapes.

Proper management of these primary pests is the most important consideration in an IPM program. Grape pests and the damage they cause needs to be recognized so that populations can be monitored at the appropriate time of the season to determine if insecticide applications are warranted. Avoiding unnecessary sprays will help prevent outbreaks of secondary pests that are normally regulated by an assortment of predators and parasites.

**Leafhoppers**

*Virginia creeper leafhopper, Erythroneura ziczac Walsh*  
*Western grape leafhopper, E. elegantula Osborn*

In addition to the widespread Virginia creeper leafhopper (VCLH), the western grape leafhopper (WGLH) that was initially found infesting grapes on the east side of the Okanagan Valley from the north end of Penticton south to the U.S. border has now spread to the west side of the valley and as far north as Naramata and Peachland. The WGLH is more tolerant to insecticides and it is important to determine if it is present in your vineyard. Leafhoppers are often distributed unevenly in vineyards and monitoring will help determine which areas require treatment. Unnecessary sprays reduce numbers of beneficial insects and spiders, resulting in a rapid resurgence in pest numbers and outbreaks of thrips, mites, and mealybugs.

Experience has shown that WGLH numbers are low in organic vineyards; damaging populations in conventional vineyards generally result from repeated sprays of insecticides that do not control this species but damage numbers of the egg parasite, *Anagrus erythroneura*. Successful control of leafhoppers relies on proper monitoring of numbers throughout the vineyard and the judicious use of insecticides in order to preserve beneficial insects and spiders.

**Identification**

Adults of both species are similar in appearance, nearly ½ cm in length with reddish-brown markings on a pale white or yellowish background. Adult WGLH (2d in photographic guide) can be distinguished by their pale eyes and irregular reddish-orange markings on a whitish background. VCLH (2a) have reddish-brown eyes and a more regular reddish-brown zigzag pattern on the wings. The background body colour is pale yellow to light brownish-yellow, producing an overall darker appearance. Adult leafhoppers are best identified on yellow sticky cards placed in outer vineyard rows during late April and May that can be inspected with the aid of a hand lens or magnifying glass.

Immature WGLH (nymphs) (2e) are distinguished by their pale white colour, lack of pigmentation in the eyes, and the presence of one to three pairs of pale indistinct yellow spots on the thoracic segments of larger nymphs. VCLH nymphs (2b) have reddish-brown eyes and a pale yellowish body colour. Larger VCLH nymphs develop a pair of dark reddish-brown spots on the first body segment behind the head and a pair of reddish-orange spots on the other two thoracic segments. Nymphs can be identified feeding on the undersides of fully expanded leaves that show signs of feeding damage (stippling).
Life Cycle and Damage

The biology and life cycles of the two species are similar. There are two generations each year. Adults of both species spend the winter in leaf litter or under plant debris in the vineyard or in nearby vegetation. Adults emerge on warm spring days to feed on a wide variety of plants, moving to grape, Virginia creeper and Boston ivy when the first leaves appear. The overwintered females deposit eggs in the leaf tissue on the undersides of fully expanded leaves during May and June. The small, flattened opaque eggs (2c) are deposited side by side, usually in small groups of around two to six for the VCLH and singly for the WGLH. Nymphs of this 1st generation hatch mainly from mid-June to the end of July. There are five nymphal stages. Winged adults that appear during July and August lay eggs that develop throughout the fall into overwintering adults. The WGLH develops more slowly than the VCLH and nymphs will occur later into the year.

Both adults and nymphs feed by piercing individual leaf cells and sucking out the contents. Light infestations cause leaves to appear stippled (2f) due to the death of individual cells. Heavier feeding results in brown, dried leaves that fall prematurely. Some studies have shown that light infestations can actually improve the quality of grapes on vines that are overly vigorous, but excessive feeding reduces the photosynthetic activity of the vines, resulting in delayed maturity, yield losses, and reduced fruit quality. Large infestations deplete carbohydrate stores and weaken the vines. Light coloured varieties can become spotted and unsightly with excrement, which is a particular concern for table grapes (2g). Adult leafhoppers are also an annoyance to pickers during harvest.

Monitoring and Spray Thresholds

Overwintered adults usually congregate in higher numbers on the edges of vineyards or in sheltered locations, and these areas should receive particular attention in the spring. Leafhoppers prefer vigorous vines and those that leaf out earliest. Monitoring numbers of adults captured in spring on yellow sticky cards can help determine relative infestation levels, but because some eggs fail to develop and predation rates are variable, these numbers do not correlate well with subsequent nymph counts. It is helpful to note where large numbers of adults occur early in the season, as these areas should be observed more closely later on.

Adults are more tolerant of insecticides and sprays should be targeted against the wingless immature stages. Depending on the location of the vineyard, monitoring of 1st generation nymphs should begin in early to mid-June. Monitor populations by counting numbers of nymphs on the lower surfaces of 5-10 leaves in at least 5 locations per block or variety. Select older bottom leaves up to just above the fruiting zone. In order to prevent damage to any vines, sampling should focus on more heavily infested areas. Sample from the north and east sides of rows where more nymphs will be found and focus on the leaf zone showing signs of feeding damage. Infestation levels are based on the average number of nymphs per leaf.

Established grapevines are tolerant of leafhopper damage during the first half of the season when growth is rapid. Because of this, there are no established thresholds for 1st generation nymphs. The threshold of approximately 20-25 nymphs per leaf established for the 2nd generation in late summer can also be used as an approximate threshold for early summer sprays. A higher threshold can be used for 1st generation nymphs if healthy populations of predators are present and a high rate of parasitism is likely. Control of 1st generation nymphs is often more effective as there is less foliage, nymphs are confined to leaves around the fruiting zone, and sprays can be directed in a narrow band to ensure good coverage. It is recommended, therefore, to treat vineyards with a history of heavy leafhopper damage during this time. Soap sprays are much more effective when targeted against 1st generation nymphs when approximately 80% of the eggs have hatched. Egg hatch can only be determined with the aid of a low power microscope, but the presence of mostly small, young nymphs can be used as an approximate measure. Because soaps are not as effective as other insecticides, the threshold for vineyards that are managed organically should be reduced considerably, perhaps to around 5-10 nymphs per leaf.

Second generation nymphs should be monitored beginning in August in a manner similar to that outlined above. Greater attention should
be paid to vines in the center of the field, however, as infestations will have spread from field edges and ‘hot spots’ to a larger area of the vineyard. Nymph counts should be collected initially every other week and then weekly when the time for spraying approaches. For wine grapes, an approximate threshold of 20-25 nymphs per leaf on the more heavily infested leaves can be used as a rough guideline. This is an average value based on counts from several leaves as outlined above. Again, for a particular area of the vineyard, collect 5-10 leaves from at least 5 areas and calculate the average number of nymphs per leaf. For 2nd generation nymphs, sample leaves from the middle of the canopy above the fruiting zone and from the north or east sides of vines where numbers will be higher. Leafhoppers will be distributed unevenly in a vineyard and it is important to sample thoroughly to determine which areas require treatment. Although the above threshold provides a rough guideline, an acceptable spray threshold should be determined by individual growers based on their past experience and additional factors such as vine vigour and numbers of beneficial insects.

**Biological Control**

Predators, such as birds and spiders, and parasites attack leafhopper eggs, nymphs or adults. Although occasionally contaminating table grapes, earwigs consume a large number of leafhopper eggs, leaving behind small, shallow scraped areas on the undersides of leaves. A small egg parasite, Anagrus daanei, helps control VCLH in some vineyards where parasitism rates can approach nearly 100% late in the season. A different parasite, A. erythroneurae, parasitizes eggs of the WGLH and is largely responsible for the control of this pest. A. erythroneurae overwinters in eggs of the rose leafhopper on wild and domestic roses, apple, blackberry and plum. In early spring this parasite utilizes eggs of a leafhopper found on a wide range of mints, including catnip and catmint. Most vineyards have good numbers of A. erythroneurae and outbreaks of WGLH are largely associated with sprays of insecticides that do not control this leafhopper but are toxic to the parasite. Excessive rates or unnecessary sprays of insecticides will also reduce numbers of other beneficial insects and spiders.

**5 Crop Protection**

**Cultural and Physical Control**

Leafhoppers prefer excessively vigorous plants and vines should be irrigated and fertilized to maintain moderate growth best suited to the production of high quality wine grapes. The use of yellow sticky tape applied below the cordon in spring can be an economical way to manage leafhoppers in some vineyards. Although costly, this method helps preserve beneficial insects and reduces or eliminates the need for additional insecticides later in the season. The use of sticky tape is likely not cost effective for large areas but it can be a useful method where infestations are confined to small areas or to the edges of vineyards.

A study conducted by PARC researchers beginning in 2001 demonstrated that removal of basal leaves during the 2nd to 3rd weeks of June when most eggs of the 1st generation have been laid effectively reduced numbers of leafhoppers and the incidence of bunch rots. No significant differences were noted in fruit quality other than a slight reduction in yield and berry size, but it is recommended that growers first assess the effectiveness of early leaf removal on only a few rows of each variety. Removal of leaves around the fruiting zone at any time of the season might not be suitable for stressed vines or on sites with intense heat and light that might lead to sunburn. However, sunburn is less likely to occur following early removal of leaves (June) as compared to late (August) leaf stripping. Exposure to more light and air causes developing berries to develop thicker skins and more wax, and the subsequent growth of lateral shoots will provide some shading. This practice is labour intensive but growers often remove these leaves later in the season in order to improve fruit quality. Removing basal leaves too early will simply force leafhoppers to deposit their eggs higher up the vine and might inhibit bud initiation required for the following year.

**Chemical Control**

Some insecticides registered for leafhopper control are ineffective for the WGLH and it is important to determine if this species is present in your vineyard. If only the VCLH is present, natural enemies should be preserved by applying the lowest effective rate only to areas where the pest has exceeded the threshold level. To date,
this species has not yet developed resistance to insecticides.

When monitoring of leafhopper nymphs indicates that insecticide sprays are required, apply one of the materials listed in Pest Control Products Recommended for Use on Grapes in British Columbia, posted at http://www.agf.gov.bc.ca/cropprot/grapeipm/grape_pesticides.pdf.

**It is often sufficient to spray small areas or treat only the perimeter of the vineyard.** Spraying in this fashion is more economical and helps preserve beneficial insects. Although a reduced rate (640 ml/ha) of carbaryl (Sevin XLR Plus™) to control VCLH is suggested to help preserve beneficial insects, full label rates will provide only moderate control of the more resistant WGLH. Malathion is currently registered for the control of leafhoppers on grapes but it is largely ineffective against the WGLH. Assail™ (acetamiprid) has been shown to be repellent to adults and will cause them to move to unsprayed portions of the vineyard. Sprays should be targeted against small nymphs. Although Assail™ is less damaging to some beneficial insects compared with many broad-spectrum insecticides, it is toxic to predacious thrips and can cause spider mite numbers to increase. Leafhopper numbers will be suppressed with sprays of Surround™ (kaolin clay) or from sprays of PureSpray Green™ spray oil or Safer’s Insecticidal Soap™ applied for the control of mites and grape mealybug. Apply soap and oil in high volume sprays and ensure that the bottom leaf surfaces are covered thoroughly.

Recent research at AAFC-PARC Summerland has shown that some newer fungicides and surfactants are repellent to adult leafhoppers and their use might eliminate the need for additional sprays. Increased movement of adult leafhoppers within and between vineyards requires greater vigilance.

**Sharpshooters**

The poplar sharpshooter, Neokolla heiroglyphica (Say), and the willow (=purple) sharpshooter, N. confluens (Uhler), occasionally infest the shoots of grapevines in sufficient number to warrant control. The dark coloured adults appear oversized in relation to the other leafhoppers found on grapes in British Columbia and they have more pronounced triangular shaped heads (2h). The large nymphs appear a uniform brown. Infestations are generally restricted to vines growing in close proximity to the preferred host plants after which these species are named. There are no chemicals registered for the control of sharpshooters in British Columbia, but sprays applied for the control of leafhoppers should also help control these species.

**Climbing Cutworms**

*Lepidoptera: Noctuidae*

To date, over 20 species of cutworm have been identified feeding on the buds of grapevines in the southern interior of British Columbia; the three most common species being Barnes’ (=well-marked) climbing cutworm, Abagrotis orbis (Grote), Reed’s dart, A. reedi Buckett, and Abagrotis nefascia (Smith). The W-marked cutworm, Spaelotis clandestina, old man dart, Abagrotis vetusta, and dark-sided cutworm, Euxoa messoria, can also cause considerable damage in certain locations in some years and there is growing concern about a new invasive cutworm pest, the lesser yellow underwing, Noctua cenea, that continues to increase in numbers and distribution since it was first found in the Okanagan in 2001. Cutworm feeding damage to the buds of grapes can vary greatly between years and locations and it is not uncommon for only parts of vineyards to be seriously damaged. Because populations fluctuate greatly and monitoring of cutworm larvae is difficult, damage can be considerable.

**Identification**

Cutworm larvae are smooth, soft, thick-bodied caterpillars measuring about 4 to 5 cm when fully grown (1a, 1b). Larvae typically live in the soil or under debris and will curl up when disturbed or handled. Depending on the species, larvae are most often light to dark grey or brown and marked with spots and stripes. Adults are drab moths with bodies around 2 to 2.5 cm long (1a, insert). The hind wings, which are sometimes distinctly marked, are usually lighter in colour than the forewings. The lesser
yellow underwing moth is most noticeable by the bright orange hind wings that are edged in black (1b). Cutworm moths are active at night and rest during the day in sheltered locations.

**Life Cycle and Damage**

Cutworm that feed on grapes spend the winter as small larvae in the soil or under fallen leaves or debris. When the weather warms in spring they resume feeding and complete their development. Damage to grapes occurs during this time when the rapidly growing larvae feed on developing buds and new shoots. Moths emerge in June and remain in a quiescent state throughout the summer until fall when they become sexually mature and begin laying eggs that hatch into the overwintering larvae. There is a single generation each year.

Damage to grape buds varies with the size of the cutworm larvae. Small larvae will often burrow into a bud, creating nearly circular holes that were previously attributed to feeding by adult click beetles (1c). Larger cutworm larvae will chew large, sometimes ragged, holes and remove a large portion or all of the bud. Feeding on new, tender shoots can cause the shoots to break or wilt (1d). Larvae climb the vine trunks or support posts each night and return to the soil during the day, or they will find refuge under loose bark of older vines.

**Monitoring and Spray Thresholds**

Cutworm numbers vary over time and between and often within vineyards. Some vineyards rarely experience significant levels of cutworm feeding damage while others are damaged yearly. Mild, dry conditions during the winter increase survival rates, while outbreaks of viral and other diseases that affect cutworms can reduce numbers to low levels. Cutworm damage is often higher in areas with dry gravelly or sandy soils that do not support good stands of broadleaf weeds, with drip irrigation systems, or as a result of weeds being controlled too early in the spring before shoots have elongated. Maintaining a record of previous damage can help suggest which areas need to be monitored most closely and where damage is likely to occur first, but bud damage needs to be assessed throughout the vineyard.

New vines planted in untreated land previously in pasture are prone to damage and larvae are often sheltered and somewhat confined within the protective milk cartons or sleeves placed around new vines. Loss of buds on new vines can retard growth and delay fruiting and they should be closely inspected in spring for signs of damage and treated when necessary.

Significant amounts of damage can occur over a period of two or three days when the nights remain warm and it is especially important to monitor for damaged buds frequently during this time. At other times it is usually sufficient to monitor vines weekly or twice weekly until shoots have elongated. An accurate threshold based on the degree of bud damage is difficult to determine, as it is influenced by the variety, training system, method of pruning, presence of predators and parasites, and tolerance level of the individual vineyard manager.

Suggested economic thresholds provided by growers range from virtually zero to over 15%; a value around 3% bud loss is likely for most vinifera varieties. Establishment of spray thresholds should also consider the potential for secondary pest outbreaks that will require additional insecticide sprays later in the season.

**Biological Control**

Cutworm eggs are consumed by various predators and are parasitized by small wasps. Larvae are eaten by ground beetles, hunting spiders and birds, and are parasitized by wasps and flies. Fungal and bacterial diseases that attack cutworm larvae are common, particularly when conditions are cool and wet. The combined effect of these natural controls is to eliminate most cutworm eggs and larvae. Of the 300 or more eggs laid by a female moth, only a few survive to become reproductive adults. Even so, damage resulting from the few remaining larvae can be considerable.

**Cultural Control**

Broadleaf weeds growing in the vine rows should not be controlled in spring until shoots have elongated and the first leaves have expanded. Damage is generally much lower in vineyards with mixed ground cover vegetation and where broadleaf weeds, particularly winter annual mustards such as shepherd’s purse, Capsella bursa-pastoris, are present in the
vine rows and drive rows in spring. Larvae are usually more common in areas with well-drained sandy or sandy-loam soils that support fewer broadleaf weeds. Diseases that infect larvae are also likely to be more prevalent in soils that retain moisture. Control of vegetation with herbicides or repeated discing during the fall prior to the establishment of a new vineyard will help reduce damage to newly-planted vines.

Cutworm damage is partially related to pruning practices and grape varieties. Vines with a greater number of retained buds in spring will suffer marginally less damage, but there will be added costs related to removal of extra shoots. Some grape varieties will produce fruit on secondary shoots, but yields will still be reduced and ripening can be delayed or uneven.

**Chemical Control**

Sprays of the pyrethroid insecticide Pounce™ (permethrin) should be directed to the soil at the base of vines, to vine trunks, and to support posts so that larvae contact the insecticide while moving from the soil to the canopy. Do not cultivate or disturb the soil for several days after treatment. For young vines, sprays should be directed into the milk cartons or other protective structures applied around newly planted vines. Avoid spraying insecticides unnecessarily, as these are very damaging to beneficial insects and can result in outbreaks of secondary pests later in the season.

Altacor™ (rynaxypyr) was recently registered for cutworms on grapes. Rapid cessation of feeding and toxicity result largely through ingestion and sprays should be directed to the developing buds. Altacor™ is considered a reduced-risk material due to its greater selectivity and reduced toxicity to non-target organisms. Follow the label instructions and wear appropriate protective clothing as for all pesticides. The list of materials registered for cutworm control is likely to change before the next printing of this guide and it is important to refer to the most recent edition of Pest Control Products Recommended for Use on Grapes in British Columbia for appropriate registered materials.

**Grape Phylloxera**

*Daktulosphaira vitifoliae* (Fitch)

Grape phylloxera (*Homoptera: Phylloxeridae*), native to eastern North America, is a type of aphid found on the roots of grapes or in galls formed on the leaves of wild grapes or hybrid varieties. European *vinifera* varieties are particularly susceptible to this pest and the accidental introduction of phylloxera to California, Europe, and elsewhere initially devastated these industries. Management of phylloxera with insecticides was not successful and the wine grape industry was only rescued with the development of resistant rootstocks derived from American *Vitis* species or hybrids between these and susceptible *V. vinifera*. Phylloxera were first identified in the Penticton area in 1961; infestations currently are scattered and at relatively low levels in British Columbia vineyards, mostly due to the use of resistant rootstocks and the failure of this pest to thrive on sandy soils.

As early as 1914 it was recognized that grape phylloxera consisted of several distinct biotypes or races. Biotypes differ in their ability to form galls on the leaves of certain cultivars, in the degree of feeding damage caused to roots, and their propensity to form sexual or winged forms. One study demonstrated that rootstocks with unsatisfactory levels of resistance to phylloxera in Europe and South Africa were suitable for California, Australia and New Zealand. Loss of resistance can also occur over time as new biotypes of phylloxera arise.

**Identification**

Adults are very small, usually wingless, and less than 1mm long and half as wide (4a). Colour varies from pale green, yellowish-green or light brown on vigorous roots, to brown or orange-brown on declining roots; as adults age they become a darker brown or even purplish-brown. Winged sexual forms are rare in British Columbia and the northwestern United States and they do not appear able to reproduce sexually on *vinifera* grapes. Leaf galls are formed on French-American hybrids, Concord juice grapes and some rootstocks. The relatively small galls on the undersides of leaves are green and roughly circular (4c). These should not be confused with the white or brown fuzzy galls of variable sizes and shapes formed on the under-
sides of leaves by erineum (leaf blister) mite (4d).

**Life Cycle and Damage**

Phylloxera overwinters on roots as small, dark hibernating nymphs that complete development in spring to become females that reproduce without having mated. Two or three generations are produced each year. When populations are large or the vines are declining, some newly hatched nymphs leave the roots through cracks in the soil and migrate to the roots of nearby vines. These small crawlers have been known to climb vine trunks and be blown on the wind for some distance. In late summer a portion of the nymphs develop into winged females (alates) that leave the soil and fly to other vines where they lay eggs in the crevices in the bark that hatch into males and females. After mating, females produce a single larger overwintering egg. In spring the eggs hatch into females that feed on developing leaves and produce galls on susceptible varieties. Nymphs produced in the galls can infest other leaves or migrate down to the roots. Most grapes grown in British Columbia are not susceptible to leaf galls and the life cycle here largely consists of asexual forms confined to the roots of grapes. For this reason, infestations spread at a relatively slow rate.

Feeding by phylloxera on the roots of grapevines results in severe water and nutrient stress and eventual death of the vines. Roots become stunted and galled and die prematurely from the injection of harmful saliva and invasion of fungal pathogens (4b). Depending on vine vigour and the size of the infestation, vines can slowly become stressed and chlorotic, or they can decline rapidly.

**Monitoring and Spray Thresholds**

Vines that fail to thrive and become chlorotic and unproductive should be inspected for the presence of phylloxera. Galls on leaves are an obvious indication of phylloxera on hybrid varieties, but damage confined to roots of *V. vinifera* grape vines can be confused with damage from nematodes or root diseases. Carefully expose the roots and search for the typical swellings and dying roots. A hand lens or dissecting microscope will help verify the presence of phylloxera.

Insecticide treatments were previously not effective and there are no spray thresholds for phylloxera on roots. Hybrid cultivars that develop leaf galls are better able to tolerate or resist feeding damage to roots, but elimination of the leaf gall forms soon after they are detected might help reduce the extent of the infestation.

**Biological Control**

Biological control of phylloxera has not been studied in detail. A number of generalist predators are known to feed on phylloxera in leaf galls. In Ontario, larvae of two species of predaceous flies were commonly associated with leaf galls. Predaceous soil-dwelling carabid beetles undoubtedly feed on phylloxera infesting roots, and because they do not produce honeydew, ants are likely to feed on phylloxera rather than protect and ‘milk’ them as they do aphids. Although natural enemies are unable to effectively control populations of phylloxera, diseases and predators will help keep populations low and reduce the rate of spread.

**Cultural Control**

When establishing a new vineyard, purchase plants from reputable suppliers and ensure that planting material is healthy and free of phylloxera. Vines imported into British Columbia are given a hot water treatment to prevent the importation of phylloxera and other grapevine pests.

To ensure that phylloxera are not moved from infested to clean areas on farm machinery, disinfect all equipment. Less frequent cultivation is thought to be helpful.

**Resistant rootstocks provide an effective means of combating phylloxera.** Almost all resistant rootstocks were developed from crosses between several species of wild grapes native to eastern and southern North America (*V. riparia, V. rupestris, V. berlandieri* and others). Some of the grape rootstocks with good phylloxera resistance that are commonly grown in British Columbia include SO4, 5BB, 5C, 420A, and 161-49C. A more complete listing of phylloxera-resistant rootstocks that includes other characteristics can be found in ‘Phylloxera-Resistant Rootstocks for Grapevines’, Northwest Berry and Grape Information
Rootstocks vary in characteristics such as vigour and it is often difficult to select an appropriate rootstock for a particular vineyard. Soil types, resistance to nematodes, vine spacing and several other factors need to be considered in addition to phylloxera resistance. Prior to planting it is advisable to read appropriate publications and consult with vineyard managers or consultants.

Vigorous vines resist phylloxera damage better than stressed vines. Provision of additional water and nutrients can help delay the rate of decline. Infested vines should not be heavily cropped and extra care is required to prevent additional stresses from insects and diseases.

Some varieties are more vigorous and are better able to resist damage. Phylloxera infestations are more damaging and spread more rapidly on heavier soils that are prone to cracking, while vines planted in sandy soils can be nearly immune to attack.

Chemical Control

Movento™ (spirotetramat), a systemic insecticide effective against plant-feeding insects with sucking mouthparts, was recently registered for the control of grape phylloxera and mealybug. Sprays should be applied in a minimum spray volume of 500 L/ha as per the label instructions. Do not apply Movento to table grapes.

Wasps

A number of species of yellowjacket wasps (Hymenoptera: Vespidae) are pests of grape; the most numerous and troublesome being Vespa pennsylvania (Saussure). Two uncommon species of paper wasp (Polistes sp.) are native to our area. The European paper wasp, P. dominula, was found in British Columbia in 2003 and has since become very common, particularly around buildings. Yellowjackets, hornets and paper wasps are important predators of grape pests throughout most of the year, but in fall they often feed in large numbers on damaged fruit. Wasps are also an annoyance to vineyard workers and to pickers during harvest operations. Paper wasps are less aggressive than yellowjackets or hornets and are not as prone to sting.

Identification

Yellowjacket workers are about 1.5 cm in length; the abdomen appears largely yellow with thin black bands. Hornets are slightly larger and appear largely black with yellow or white bands. Paper wasps have thinner bodies with a more pronounced constriction (stalk) between the thorax and abdomen. The body is dark with some yellow markings, but they appear reddish-brown due to the colour of the wings and distal parts of the legs (5h). In flight the legs of paper wasps dangle visibly below the body. All of these wasps produce paper nests, but the brood cells of paper wasps are exposed.

Life Cycle and Damage

Yellowjackets and paper wasps overwinter as fertilized females in sheltered locations. These queens seek suitable nest sites in spring and establish a new colony. Nests are initially small, consisting of only a few paper cells, but as workers develop they assist in building up colony numbers and the size of the nest. Depending on the species, nests can be in the ground, under the eaves of houses, in hollow trees, the walls of buildings, or exposed in trees. In fall, in addition to workers, colonies produce both males (drones) and queens. When freezing weather arrives, all the drones and workers, as well as the undeveloped larvae and pupae are killed.

Wasps prey on other insects, scavenge food and feed on nectar and the juices of fruit. It has been reported that wasps do not damage fruit themselves but take advantage of injury caused by other insects or birds, diseases such as sour rot, or splitting of fruit. There are contrary reports, however, particularly relating to the European paper wasp, that they are able to directly feed on intact fruit (5h). As mentioned above, wasps are also a major annoyance and their stings result in swelling and a painful itch and can trigger severe allergic reactions in some individuals.
**Monitoring and Spray Thresholds**

There are no established thresholds for wasps and the need to spray is determined by the variety of grape, past history of damage, and tolerance level of individual vineyard managers. Trapping of wasps is most effective early in the season before the colonies have increased in number. The recent arrival of the European paper wasp has made it necessary to control wasps more frequently.

**Biological Control**

Wasp colonies can be attacked by other wasps. Eastern and western kingbirds feed on foraging adults and larvae are subject to certain diseases. The onset of winter eliminates established colonies and overwintering queens often succumb to cold or are eaten by other insects, or even by mice.

**Cultural Control**

Elimination of nests early in the season can be an effective management strategy, but they are often located outside the vineyard or are well hidden. Aerial nests in trees or under eaves can be removed and placed in a plastic bag at night when wasps are inactive. Nests in walls or natural cavities can be treated by spraying or puffing commercial wasp control products into the entrance holes at night. Sealing of entrances can help prevent future nests.

Trapping can be used to reduce numbers of foraging wasps in localized areas. Commercial wasp traps that use an attractant dissolved in water (e.g. Rescue® Yellowjacket trap) have proven effective, or homemade versions containing a sugary solution can be made from recycled 1 litre plastic pop bottles. Wasps enter through small holes and eventually fall into the liquid solution and drown. Trapping early in the season when populations are low and queens are establishing colonies is generally more effective than attempting to remove large numbers of wasps later in the season. For additional information on social wasps, management strategies, and design of wasp traps, visit the University of California Statewide Integrated Pest Management Program, 2008, website at: http://www.ipm.ucdavis.edu/PMG/ PestNotes/pn7450.html.

**Chemical Control**

Cypermethrin (Ripcord™) is currently registered for the control of wasps feeding on wine grapes but not on table grapes. The pre-harvest interval (PHI) is two days for wine grapes harvested by machine and seven days for fruit harvested by hand.

**Spotted Wing Drosophila**

*Drosophila suzukii* (Matsumura)

This potentially damaging pest of grapes was first found in Canada infesting blueberries in the Fraser Valley in September 2009 and trapped soon after in a cherry orchard near Kelowna. It is native to Southeast Asia where it feeds on a range of soft skinned fruit. Unlike most species of fruit fly (vinegar fly) that feed on very ripe, damaged, or rotting fruit, the spotted wing drosophila (SWD) is able to attack ripening, firm fruit.

**Identification**

Adult spotted wing fruit flies are slightly larger (2-3 mm) than other fruit flies and males have a dark grey to black spot toward the outer margin of each wing (5g). Females lack the wing spots and have a saw-like ovipositor visible with the aid of a good hand lens or microscope. The elongate creamy white legless larvae, up to 3 mm long, are difficult to distinguish from other species of fruit fly.

**Life Cycle and Damage**

SWD overwinter as adults. Beginning in spring, several generations develop throughout the year on a range of available fruit (strawberry, blackberry, cherry, etc.), including grapes in the late summer and fall. Typical for fruit flies, females lay large numbers of eggs that hatch rapidly into larvae that mature at a rapid rate. A generation can develop in as little as 1 to 2 weeks when temperatures are warm, resulting in rapid increases in numbers and significant damage to crops.

Larvae of several species of fruit fly develop in damaged or decaying grapes late in the season, causing little damage to the crop. Female SWD are able to deposit eggs into undamaged, ripening table grapes with the use of their strong
serrated ovipositors and it is possible that a small proportion of thin-skinned wine grapes are damaged in this same manner. Recently completed studies suggest that SWD causes little direct economic damage to wine grapes grown in south central British Columbia. Laboratory studies have shown that larvae were unable to develop in wine grapes until fruit had reached a maturity level of nearly 20 °Brix. It was not collected from intact, healthy fruit and formed a smaller proportion of all Drosophila species found in damaged or rotting grapes. SWD was found, however, infesting apparently healthy table grapes. The presence of fruit fly larvae and pupae in fresh table grapes is a major concern. Feeding by larvae directly damages the fruit and the holes created during egg-laying allow the entry of disease-causing microorganisms. The relationship of SWD and other drosophila to the spread of sour rot and other pathogens affecting ripening grapes is not entirely clear.

**Monitoring and Spray Thresholds**

There are no established thresholds for this pest and it remains unclear if it poses a serious threat to wine grapes produced in British Columbia. Fruit fly traps and baits are available commercially or they can be made from recycled plastic pop bottles containing mixtures of yeast, sugar and water or apple cider vinegar as bait. Fruit can also be carefully inspected for the presence of larvae or pupae and adults reared out from infested fruit in order to provide an accurate identification. If you think you have this pest, consult the ‘Spotted Wing Drosophila (Fruit Fly) Pest Alert’ factsheet, BCMA (http://www.agf.gov.bc.ca) for additional information.

**Biological Control**

Parasitic wasps have been reared from SWD in BC and predators of other species of fruit fly are likely to also attack this species.

**Cultural Control**

Sanitary measures suggested for other fruit crops (i.e. removal of infested and unharvested fruit) are generally not considered to be practical for commercially produced wine grapes. Early season removal of basal leaves (see leafhopper section for details) might help control this and other fruit fly pests. Exposure of fruit early in the season results in fruit with thicker skins and heavier wax cuticles which helps control sour and bunch rots.

**Chemical Control**

Emergency registration of Ripcord™ (cypermethrin), Malathion, Delegate™ (spinetoram), and Entrust™ (spinosad) for the control of SWD has been requested for 2014. Entrust is a product approved by the Organic Materials Review Institute (OMRI) for use in organic vineyards. Sprays are required near to harvest to control this pest and it is important to consult the label and observe the pre-harvest interval (PHI). Sour and bunch rots that might be spread to some degree by SWD are best managed through the combined use of early season removal of basal leaves and judicious use of fungicide sprays.

**Grape Mealybug**

_Pseudococcus maritimus_ (Ehrhorn)

Grape mealybug was previously not considered to be major pests of grapevines grown in British Columbia, as numbers were generally quite low due to the activity of parasites and predators. The recent recognition that they and soft scale are responsible for the sometimes rapid spread of grapevine leafroll associated virus 3 has raised its status to that of a major pest. Direct feeding damage is of little concern. Individual bunches of table grapes touching infested vines would occasionally become infested and fruit can become coated with sticky honeydew, reducing their marketability. **Largely due to the detrimental effects of broad-spectrum insecticides on beneficial insects, damaging populations of mealybug have become more common in recent years.** Whenever possible, apply chemicals against major pests (leafhoppers, cutworm) of grapes only when required and only to portions of the vineyard where control is warranted. Select pesticides that are least damaging to predators and parasites.

**Identification**

The grape mealybug (_Homoptera: Pseudococcidae_) has a flattened, oval pink body covered in a mealy white wax coating (3f). It is somewhat segmented in appearance, but the divisions
between head, thorax and abdomen are not distinct. Mature wingless females are about 5 mm long. They have long waxy filaments along the edge of the body that are longest at the rear and become progressively shorter toward the front end. Large numbers of eggs are laid in cottony masses. The smaller, winged males have a pair of long, white anal filaments. Yellow to brown crawlers that emerge from the oval, orange eggs are not covered in wax. All stages are mobile.

**Life Cycle and Damage**

Females move in late fall to old wood and lay overwintering eggs in cottony egg masses under loose bark. Crawlers, some of which might have hatched in fall and remained dormant throughout the winter, move to new shoots in spring. Grape mealybugs mature around mid-summer and produce a 2nd generation that matures in late autumn. They can occur on all aerial parts of the vines but are more common in summer on leaves and new growth, usually hidden within the canopy.

Damage is similar to that for soft scale. Plant sap is removed during feeding and large amounts of honeydew foul the fruit (3f) and promote the growth of sooty mold fungus. Most fruit damage occurs during the development of the second brood in late summer. Mealybugs can infest grape bunches and are known to transmit certain leafroll viruses.

**Monitoring and Spray Thresholds**

To detect when crawlers are active, remove the loose bark in an area near the infestation and encircle the vine with clear tape applied sticky side out, or use double sided adhesive. Replace the tape every three or four days and check for the yellowish brown crawlers. A good magnifying glass or dissecting microscope is useful for this purpose. Later in the season inspect vines for the cottony egg masses, leaves coated with shiny, sticky honeydew, and the presence of attendant ants travelling up and down the vine trunks. The need to inspect vineyards for mealybugs can be based partly on past infestations, vigorous growth, and prior use of broad-spectrum insecticides. **Grape mealybug prefer vigorous vines with thick canopies.**

**Biological Control**

Parasitism of grape mealybug has not been studied in British Columbia. Research in California has shown that at least six species of parasitic wasps attack them - late summer and fall rates of parasitism often exceed 90%. Predaceous midge larvae feed on eggs, while several species of lady beetles and many other generalist predators feed on nymphs and adults. Mealybug and other homopteran insects (scale, whitefly, aphids, etc.) are susceptible to several diseases, including *Entomophthora* fungus.

**Cultural Control**

Mealybugs prefer vigorous vines. Ensure adequate nutrition and moisture to produce an optimum crop, but avoid excessive vigour. Research in California has shown that fruit on cane-pruned varieties is less likely to suffer damage compared with spur-pruned vines. Clusters on cane-pruned vines hang farther away from the mealybug overwintering sites on old wood. Earlier varieties are not exposed to honeydew to the same extent as later varieties.

Heavy infestations of mealybugs and other honeydew-producing pests of grape are usually tended by ants that feed on the sugary excretion and protect the mealybugs from predation. Control of ants can often contribute to a significant decline in mealybug numbers. For small backyard plantings with an isolated infestation, painting a sticky material around vine trunks and posts and ensuring that ants cannot gain access to the mealybugs often eliminates the problem. Trials in California have also shown that sprays directed against the Argentine ant, *Iridomyrmex humilis*, controlled damaging populations of the obscure mealybug, *Pseudococcus affinis*. Insecticides are not currently registered for the control of ants in BC vineyards.

**Chemical Control**

Malathion, Diazinon, and Safer’s Insecticidal Soap™ can be applied against the crawler stage, while Movento™ (spirotetramat) is active against all stages due to its good systemic activity. Movento™ should not be applied to table grapes.
Soft Scale

European Fruit Lecanium Scale,  
Parthenolecanium corni (Bouché)  
Cottyn Maple Scale, Neopulvinaria innumerabilis  
(Rathvon)

About a dozen species of scale insects are known to attack grapes in various regions of the world, but only two are commonly found on grapes in British Columbia. The European fruit lecanium scale (EFLS) and cottyn maple scale (CMS) are both soft scales (Homoptera: Coc-cidae) that rarely cause direct economic injury to grapes. They are a greater concern in the production of table grapes. The large quantities of honeydew they produce supports the growth of sooty mold fungus that can leave fruit unmarketable. Two species of hard scale (Homoptera: Diaspididae), the oystershell scale, Lepidosaphes ulmi L., and San Jose scale, Quadraspidiotus perniciosus Comstock, can occasionally be found attacking 2 year old wood of weakened vines. Hard scales do not produce large amounts of honeydew and are currently of little importance to the British Columbia grape industry. CMS and EFLS are considered major pests of grapevines due to the recent discovery that they are effective vectors of grapevine leafroll virus 3.

Identification

Female EFLS, also known as brown apricot scale, when fully grown have a chestnut brown, smooth and slightly shiny protective shell that is around 5 to 8 mm long and slightly longer than wide (3c). Certain races in some regions reportedly produce male scales, but most females reproduce without mating. The small nymphs or crawlers that hatch from the small oval, pearly white eggs are yellow to orange in colour. Mature female CMS are similar in size, shape and colour to EFLS, except that a mass of cottony material containing the eggs, up to 1,000 per sac, extrudes from the rear end (3d). The much smaller winged males produced in late summer have no mouthparts and live for only 1 or 2 days. Males mate with the immature females. Mature scales of both species become brittle after the eggs are laid in mid-summer and can easily be detached from the one to three year old wood with a fingernail. The smaller hard scales (1-2 mm), which are much longer than wide, do not produce any cottony material and remain firmly attached to the canes even after death.

Life Cycle and Damage

EFLS and CMS overwinter as partly grown scales on new canes (3c). Growth resumes in spring and females reach maximum size around early summer when they begin to lay eggs. The newly hatched scale called crawlers hatch and disperse during July and August to the undersides of leaves where they settle and feed until early fall, when they migrate to the current year’s wood and develop a hardened protective shell for the winter.

Both species of soft scale feed on the phloem of grapes and a large number of other woody plants, including many fruit trees and ornamentals. Soft scales have become more abundant recently. Direct feeding damage from the removal of plant fluids is rarely a cause for concern, but the additional stress caused to weak vines might be sufficient to hasten death. Of greater concern, they produce vast amounts of honeydew that fouls the fruit and supports the growth of sooty mold fungus, potentially rendering table grapes unmarketable.

Monitoring and Spray Thresholds

Partly-grown scales can often be observed during pruning, or watch for honeydew on leaves and fruit during spring and early summer. Ants feed on the honeydew and will protect the scale from predators (3d). They can often be seen in large numbers hurriedly running up and down posts or vine trunks to and from infested canes.

To detect when small crawlers are active, remove the loose bark in an area near the infestation and encircle the vine with clear tape applied sticky side out or use double sided adhesive. Replace the tape weekly and check for the orange coloured crawlers. A good magnifying glass or dissecting microscope is useful for this purpose.

There are no established thresholds for direct damage from soft scale infestations. Small to moderate numbers of scale on a healthy vine are unlikely to result in direct feeding damage due to the removal of fluids and nutrients; treatment of table grapes is required when the honeydew they excrete fouls the fruit. It is important to
control infestations of soft scale if vines or neighbouring vines are known to be infected with Grapevine leafroll virus 3.

**Biological Control**

Biological control of scale on grapes in British Columbia has not been investigated in any detail, but on other crops and in other regions of the world they are known to be heavily parasitized by several species of wasps and flies. They are preyed upon by lady beetles and other generalist predators. English sparrows and several species of warblers that feed on fully grown soft scale have been known to curtail infestations in some regions.

**Cultural Control**

Healthy vines of moderate vigour are less susceptible to scale infestations and are better able to withstand damage. Routine pruning removes many scales and a small infestation can be pruned out in spring or early summer.

**Chemical Control**

Oil or lime sulphur applied as high volume dormant sprays to the trunks and cordons of vines for the control of powdery mildew will help suppress soft scale and other small, soft bodied pests of grapevines. Movento™ (spiro-tetramat) is registered for the control of scale on wine grapes during the growing season, but it should not be applied to table grapes. Malathion can be used against thecrawler stage and insecticidal soap will provide some suppression of this active stage in summer. Repeat applications might be required as long as crawlers are active.

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### Secondary and Minor Pests

A number of insect and mite pests of grapes occur only infrequently or cause significant amounts of damage only after chemical treatments have reduced numbers of predators and parasites that normally regulate their numbers. The best way to manage these pests, then, is to properly manage populations of primary pests so as to preserve and enhance numbers of beneficial insects and mites.

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### Thrips

**Western Flower Thrips, Frankliniella occidentalis**

**Pergande**

**Grape Thrips, Drepanothrips reuteri** Uzel

**Flower Thrips, Frankliniella tritici** Fitch

Several species of thrips can be found feeding, at least occasionally, on grapes; the three most common are the western flower thrips (WFT), the grape thrips (GT), and the flower thrips (FT). **Thrips are generally minor pests of wine grapes, but are important pests of table grapes due to scarring of fruit.** Their populations often increase following applications of broad-spectrum insecticides that reduce numbers of beneficial insects. All species feed in a similar manner, but differences in biology and timing of infestations results in differential damage to grapes.

**Identification**

All species have long, slender bodies with dark protruding eyes and relatively short beaded antennae. Adults have two pairs of wings that are heavily fringed along the edges. WFT are dark yellow to brown and 1 to 1.5 mm long (5a). GT and FT are both around 1 mm long, the former is amber yellow with a yellow-orange thorax, and the latter is uniformly pale yellow to nearly white. Nymphs of all species are similar in appearance to the adults except that they are smaller, slightly paler in colour and lack wings. Thrips tend to be very active; adults not only fly but also run and hop rapidly.

Plant feeding thrips should not be confused with predaceous thrips. Adults of beneficial thrips may be dark black or purplish black with clear wings or wings that appear light grey with white bands, or they are pale with dark bands on the wings. They are slightly larger to nearly twice as large as the pest species. Nymphs of predaceous thrips have bodies that are banded yellow and orange, are reddish purple in colour, or are pale with the joints of the appendages dark. Compared with phytophagous species, predaceous thrips have front legs that are enlarged and slightly curved that they use for grasping prey. These features and the slightly larger mouthparts are best observed under a dissecting microscope.
Life Cycle and Damage

Adult female thrips overwinter under leaves and plant debris on the ground. In spring they deposit their eggs into developing leaf, stem, and flower tissues. The time from egg to adult is about three weeks during warm weather, and several generations are produced yearly. Most thrips feed on a wide range of host plants and large numbers will often move to a succession of new succulent hosts throughout the year. Both adults and nymphs feed by rasping the leaf surface and sucking up the escaping fluids.

WFT adults enter vineyards in early spring, often in large numbers, when saskatoon bushes have finished flowering and other native vegetation is beginning to dry down. In addition to ground cover plants, WFT feed on flower and fruit tissues of grapes from bloom to just after fruit set. Their feeding under caps and persistent flower parts causes berry scarring and russetting (5b). White and green grape varieties are also damaged when females insert their eggs into fruit, causing dark spots or halo spots. Feeding on developing shoots is occasionally so heavy that shoots are stunted and leaves deformed (5b), but damage of this sort often only occurs around the perimeter of vineyards. WFT usually decline greatly in numbers following fruit set. FT have a similar biology and contribute to the damage caused to fruit in spring, but they are not as numerous as WFT.

GT are generally only found in significant numbers on grape during the summer months, and their feeding is largely confined to leaves and shoots. Numbers can build up rapidly, however, particularly following insecticide treatments for other grape pests. Leaf veins can be scarred and fail to expand properly, leaf edges can become necrotic, and shoot growth inhibited, resulting in shortened internodes.

Monitoring and Spray Thresholds

WFT often invade vineyards in large numbers for only brief periods in spring and it and the FT are not normally an economic concern for wine grapes. Scarring of fruit has not been shown to affect wine quality and damage to leaves and shoots is usually minor. Damage usually occurs over a short period of time, which makes early detection and control difficult. Inspect table grapes for the presence of WFT and FT in spring from around 75% bloom to the end of fruit set and apply insecticides as required. Verify the presence of thrips by sharply striking fruit clusters or leaves against a dull white or green coloured surface. The decision to spray should be based partly on the table grape variety, records of past damage, and the intended market for the fruit. There are no firm thresholds for thrips infesting grapes in British Columbia.

Watch for GT damage to young leaves and shoots during summer and early fall. Use an eyepiece or magnifying glass to verify that thrips are present. As for fruit clusters, leaves and shoots can be held over a piece of white cardboard and sharply hit to dislodge the thrips. Control on mature vines is usually only warranted when growth is stunted sufficiently early in the summer to delay ripening and affect growth the following season.

Biological Control

Predatory bugs such as the minute pirate bug (Orius tristicolor), anthocorids (Anthocoris melanocerus), and deracorias (Deraeocoris brevis) are all known to feed on thrips. Studies in the Okanagan and Similkameen valleys have indicated that predatory thrips are important regulators of pest thrips populations. In the laboratory, predatory thrips at a density of one every 10 to 15 leaves is sufficient to control WFT.

Cultural Control

So that thrips are not forced to feed on table grapes when they are most susceptible to fruit damage, avoid mowing cover crops during bloom and fruit set.

Chemical Control

Spinosad (Success™ or Entrust™) is registered for suppression of thrips on grapes. Insecticidal soap applied for mites and grape mealybug will also help suppress thrips populations.

Spider Mites

European Red Mite, Panonychus ulmi (Koch)
Two-spotted Spider Mite, Tetramyces urticae Koch

The main species of spider mites infesting grapes in British Columbia are the European red
mite (ERM) and the two-spotted spider mite (TSSM). They differ somewhat in biology, but feeding damage and management strategies are largely the same. **Spider mites should be considered secondary or induced pests of grapes; their numbers increase following the use of broad-spectrum insecticides that reduce numbers of spider mite predators.** Although it is somewhat selective in action and preserves populations of some predators, the insecticide Assail™ is toxic to predacious thrips that are effective predators of mites in British Columbia vineyards. Studies in Washington State have also shown that the active ingredient in Assail™ and related materials increases spider mite reproduction. Due to its persistence, increases in mite numbers can occur up to several months after application. Mite outbreaks can also occur following applications of pyrethroid insecticides (e.g. permethrin) and other broad-spectrum insecticides.

**Identification**

Adults of both species are small, around 0.5 mm in length, and best viewed with the aid of a magnifying lens. They are nearly spherical in shape and have eight legs and short piercing mouthparts. The ERM is dark red in colour with long pale hairs (setae) projecting from small pale circular areas on the dorsal surface (4h). The oval eggs are bright orange with a distinctive hairlike projection on the upper side. The TSSM varies in colour from pale yellow to greenish yellow to bright orange; the common pale form has a distinct dark dorsal spot on each side of the body (4g). Its eggs are white. Both species feed on the undersides of leaves. TSSM usually congregate in clusters and produces large quantities of silk webbing, while ERM produce less webbing and are more evenly distributed over the leaf surface.

**Life Cycle and Damage**

ERM, our most common species, pass the winter as eggs laid on vine canes and trunks. Eggs hatch in spring and the young microscopic nymphs begin to feed on young foliage. TSSM overwinter as fertilized females under the bark or in other sheltered areas around the base of vines. When warmer weather arrives in spring the females begin to feed and deposit eggs. Both species can produce around six to eight overlapping generations each season and all stages can be found on grapes at any time during the summer months. Females of both species are capable of producing 200 or more eggs each and development can be rapid during warm weather. **Spider mite populations can, therefore, explode rapidly under favourable conditions.** Both adults and nymphs feed by piercing individual leaf cells and removing the fluid contents. Healthy grapevines can tolerate moderate numbers of spider mites, which will cause chlorotic spots on the leaves. Heavy feeding results in brown leaves that fall prematurely; reducing photosynthetic activity and vine vigour. Heavy feeding damage can delay ripening of fruit. The large amount of webbing produced particularly by the TSSM can also be a cosmetic problem for table grapes.

**Monitoring and Spray Thresholds**

Spider mites rarely cause significant damage to grapes prior to mid-summer. No firm thresholds have been developed for spider mites on grapes in British Columbia, but approximate guidelines can be suggested from research conducted in other countries. In Europe, a threshold for ERM on grapes was determined to be 60 to 70% infested leaves in spring, while in summer leaves with 1 or more mites should not exceed 30 to 45% of total leaves on a shoot. Because a similar threshold has been suggested for TSSM in Switzerland, numbers of both species can be combined into a single spider mite count. A good hand lens or low-power dissecting microscope would help provide accurate counts of infestation levels. A fairly reliable method is to monitor damage to leaves and spray when a moderate amount of ‘bronzing’ has occurred, around 10% defoliation. Because mite numbers can increase very rapidly
it is necessary to monitor grapes frequently during hot weather.

When monitoring for mite damage it is useful to consider previous infestation levels. Vineyards with low chemical inputs that have not experienced severe spider mite outbreaks in the past are unlikely to require treatment, while those that have been treated with broad-spectrum insecticides should be observed more closely.

**Biological Control**

Spider mites were of minor importance prior to the widespread use of synthetic pesticides. Due largely to the harmful effects of pesticides on populations of predators, spider mites are now the most important pests of grapes in many regions of the world. The first course of action for the management of spider mites is the preservation and enhancement of beneficial species. Pesticides should only be applied when necessary and only to parts of the vineyard where pest populations are sufficiently high to warrant control. Whenever possible select materials that are least damaging to non-target species. Numbers of spiders and beneficial insects are usually higher in vineyards with permanent, mixed groundcover that provides pollen and alternate sources of prey. Proximity to uncultivated or unsprayed areas allows beneficial insects to re-colonize vineyards following spray treatments.

Several species of predatory mites (e.g. *Metaseiulus*, *Typhlodromus* and *Amblyseius* species) feed on spider mites and their eggs. Most beneficial mites are a pale opaque colour but they can appear light orange if they have been feeding on ERM. They are oblong in shape and slightly smaller than adult spider mites. The long-legged orange whirligig mite (*Anystis agilis*) is uncommon on grapes in British Columbia. Like most other predatory mites, this large species is more active than its prey. Predatory mites are very sensitive to a range of pesticides, including sulphur fungicides at high rates and pyrethroids such as permethrin (*Pounce™*).

A number of spiders and predatory insects feed on spider mites or their eggs. Many species, such as the minute pirate bug (*Orius tristicolor*), are generalist predators that do not specialize on mites, while others such as the aptly named spider mite destroyer, *Stethorus picipes*, are very effective predators that feed almost exclusively on mites. *Stethorus picipes* is a small, dark species of ladybeetle with a slightly hairy appearance. The elongate bodies of the larvae are also nearly black in colour with a body covered with numerous hairs. At least four species of predatory thrips can be found in Okanagan Valley vineyards; adults of three of these are black, while the fourth, the six-spotted thrips, *Scolothrips sexmaculatus*, is pale with dark spots on the wings. All four species feed on spider mites, and healthy numbers of these predators are usually associated with low populations of their prey. The western flower thrips is considered a pest of grapes, but it is also known to feed on the eggs of spider mites.

**Cultural Control**

Vineyards can be managed in ways that help alleviate mite problems. Hot, dry and dusty conditions favour the buildup of spider mite populations. Where mites are a problem, roads can be treated with oils or other materials to reduce dust. Vineyards that are continuously cultivated will produce more dust than those with permanent ground covers. Moreover, a permanent mixed groundcover will support greater numbers of beneficial insects and predaceous mites. Compared with bare soil, planted drive rows will decrease air temperatures and raise humidity levels somewhat, providing less favourable conditions for spider mites. Spider mite populations can become elevated on vines suffering severe drought stress, and these plants are also less able to tolerate damage. Overhead irrigation will help reduce mite infestations.

**Chemical Control**

A number of miticides (e.g. *Aframite™*, *Kelthane™*, *Envidor™* and *Pyramite™*) are registered on grapes for mite control. Safer’s Insecticidal Soap™ or PureSpray Green Spray Oil™ provide a more benign approach to mite control but offer only moderate suppression of spider mites. Materials with different modes of action (chemical groups) should be used in rotation to help prevent the development of resistance.
Grape Erineum Mite
= Grape Leaf Blister Mite, *Colomerus vitis* (Pagenstecher)

**Identification**

Adult erineum mites are only 0.2 mm long and a dissecting microscope is required to observe them properly. Their creamy pale bodies are elongate with only two pairs of legs. Although erineum galls can be confused with certain fungal diseases or phylloxera leaf galls by inexperienced growers, their characteristic appearance is the best way to identify these pests. The hemispherical concave galls or erinea formed on the undersides of leaves initially have a felty white appearance due to the presence of densely packed long leaf hairs (4d). Opposite to these galls, bumps will be visible on the upper surfaces of leaves. These swellings are a reddish colour in spring and turn green as the leaves mature. Small numbers of mites feed on the dense hairs in each gall, which eventually turn brown. Two other forms of erineum mite, the leaf-curving strain and bud-inhibiting strain, have been reported recently from Washington State and are possible in British Columbia. The former causes slight to severe downward curling of leaves in summer with stunting of shoots and increased growth of lateral shoots. Feeding of the bud-inhibiting strain on buds causes a range of abnormalities, including scarified shoots, short zigzagging internodes, dead terminal buds and malformed leaves. For additional information on erineum mites and other pests of grapes, visit the University of California Integrated Viticulture Online website (Viticultural Information > Insects and Mites).

**Life Cycle and Damage**

Adult erineum mites overwinter under the scales of buds. Mites move in spring to developing shoots and create galls on young leaves around the fruiting zone. Several generations are produced each year with new galls developing higher up the shoots. Beginning in late summer, adult mites move back to the buds for winter.

Leaves with a few to many erineum galls appear to function normally and there is little or no economic damage to mature vines until almost all leaves are covered with galls. Even then, damage usually only occurs if vines are also suffering from other stresses. Establishment of newly planted vines can be delayed somewhat by erineum mite feeding, which will retard or reduce fruiting.

**Monitoring and Spray Thresholds**

Erineum galls are usually very apparent during routine vineyard operations. Grape varieties differ in susceptibility to this pest and it is important to check individual blocks for the presence of mites. In order to prevent premature loss of leaves, new vines should be inspected and treatments applied if large numbers of galls are found on most leaves during the spring or summer months.

**Biological Control**

The western predatory mite, *Galendromus occidentalis*, is an important erineum mite predator. Predaceous thrips will feed on exposed mites, but their effectiveness is likely limited when mites are protected within the dense hairs of the galls.

**Cultural Control**

New plants may become infested in the nursery. **Ensure that material used to establish new vineyards is free of mites and other pests or diseases.** Removing galled leaves can control light infestations to some extent. Research conducted in the Okanagan Valley demonstrated that removal of infested leaves in May did not alter yields or fruit quality.

**Chemical Control**

Erineum mite is seldom a problem in vineyards where sulphur is applied routinely for powdery mildew control. **Applications of sulphur (e.g. Kumulus™ DF, or wettable sulphur) are most effective early in the season when galls are first being formed on new leaves.** Sulphur sprays should not be used on Foch or *Vitis labrusca* varieties as they are susceptible to leaf and fruit damage. Miticides (acaricides) such as Pyramite™ or Kelthane™ applied against spider mites and sprays of organophosphates (e.g. malathion and Sevin™) for control of leafhoppers will also reduce numbers of erineum mite somewhat.
Grape Leaf Rust Mite
*Calepitrimerus vitis* (Nalepa)

Grape leaf rust mite, which belongs to the same family (*Eriophyidae*) as grape erineum mite, was identified for the first time on grapes in the Okanagan Valley in 2009. Growers are referred to the following source for additional information: Grapevine pests and their management, by A. Loch. Primefact 511, New South Wales Dept. of Primary Industries, NSW Australia posted at: www.dpi.nsw.gov.au/primefacts.

**Identification**

A microscope is required to properly identify these 0.2 mm long pale white wormlike mites that closely resemble erineum or blister mites, but do not form leaf galls. They are much smaller than spider mites and elongate rather than round, tapering at the rear, and having two pair of legs near the head (4e). Their presence is best determined by the bronzing of leaves during July and August that results from their feeding. Unlike spider mites that cause similar damage to leaves, grape leaf rust mites do not produce webbing.

**Life cycle and damage**

Adult rust mites that spend the winter under bark or bud scales move in spring to the developing buds to feed and lay eggs. Large numbers of adults concentrated on the available green tissue in early spring can damage buds, resulting in deformed leaves, scarring of shoots, reduced shoot growth, reduction or loss of fruit clusters, and even death of buds (4f). Early season rust mite damage can be confused with cold injury or damage from other causes. Damage becomes less visible as the season progresses and shoots lengthen. Immature mites that hatch over a period of several weeks feed under the bud scales, eventually moving to feed on developing leaves. Unlike spider mites, rust mites infest equally the upper and lower leaf surfaces. Many generations are produced throughout the summer. Before leaves drop in the fall, adult rust mites migrate to the cordon and trunk for the winter. Bronzing of leaves is most obvious during July and August, but damage to leaves is not thought to cause serious damage to the vines. Extensive bronzing of leaves is an indication that rust mites need to be controlled the following spring.

**Monitoring and Spray Thresholds**

There are no established thresholds for this pest. Control is indicated when a significant proportion of leaves become bronzed in late summer.

**Biological Control**

As for erineum mite, western predatory mite, *Galendromus occidentalis*, other mite predators, and predacious thrips are often able to keep rust mites in check. To maintain healthy populations of beneficial insects and predatory mites, it is important whenever possible to limit the use of insecticides and to apply selective materials that are less toxic to predatory mites.

**Chemical Control**

A successful rust mite control program developed in Australia relies largely on spring applications of sulphur (e.g. Kumulus™ DF, or wettable sulphur) in high volume sprays during bud swell to woolly bud stage. Sulphur sprays should not be used on Foch or *Vitis labrusca* varieties, as they are susceptible to leaf and fruit damage. There are no miticides currently registered for the control of grape leaf rust mite.

**Snailcase Bagworm**
*Apterona helix* Siebold

**Identification**

This introduced moth (Family *Psychidae*) remains throughout most of its life within a snail-like case made from particles of soil and sand bound together with silken threads. When fully grown the coiled case is approximately 4 mm in diameter (4f). There are no males and the wingless female moths remain inside the case to deposit their eggs without having mated (parthenogenetic reproduction). Large aggregations often form on posts, tree trunks, or vine trunks in late summer.
**Life Cycle and Damage**

Young caterpillars overwinter within the case formed by the mother. They drop to the ground in spring, form C-shaped cases, and begin to feed on a wide range of weeds, native plants and cultivated crops. Damage consists of small excavated holes on the leaf surface. As the larva grows it eventually expands its case into the typical snail-like, helical shape. In mid-summer the fully grown larvae will climb vines, posts or buildings, attach themselves firmly with silk, and pupate within the case. Several weeks later the females emerge, lay eggs, and then exit the case to die. The eggs hatch in late summer and the small first instar larvae spend the winter within the protective cases. There is only one generation per year.

**Monitoring and Spray Thresholds**

Larvae of the snailcase bagworm cause minimal feeding damage to leaves and they are not considered economic pests of wine grapes. The large aggregations are easily detected on buildings, posts, wires, and the trunks of vines. If contamination of table grapes is a concern, inspect vines and apply control measures before the overwintering aggregations form in late summer. The non-feeding overwintering stage, secure within the case, is not susceptible to insecticides.

**Biological Control**

Because it has not yet been adequately studied, natural enemies of this pest of grapes in British Columbia are unknown. The protective case would provide some protection from certain generalist predators, but predators, parasites and diseases undoubtedly take their toll.

**Cultural Control**

Snailcase bagworm prefer moist areas with thick vegetation. Avoid excessive irrigation and drain areas that puddle or remain damp. Bagworm are uncommon in vineyards planted on light soils with drip irrigation. Proper drainage will also help reduce grapevine root diseases. Occasional discing or mowing of areas of the vineyard where bagworm are a problem can help reduce infestations.

**Chemical Control**

Dipel™ (Bacillus thuringiensis) is registered for snailcase bagworm control on grapes and sprays applied for the control of other insects will provide some protection. For this reason, they are more common in organic vineyards or conventional vineyards with low chemical inputs.

**European Earwig**

*(Forficula auricularia* L.)*

The European earwig is both a beneficial insect and a pest of table grapes. All stages feed on small, soft-bodied insects such as small leafhopper nymphs and will scrape leafhopper eggs off of leaves. In fall they feed to some extent on damaged grapes and their presence in table grapes is not appreciated by some consumers. On grapes produced for wine they should be considered important generalist predators.

**Identification**

Adult earwigs (Order Dermaptera) are dark brown, about 1.5 cm long, with a pair of forcep-like appendages (cerci) at the rear of their elongate bodies. The anal forceps of females are long and straight, while those of males are curved. Earwigs seldom fly and the membranous hind wings usually remain folded up under the short leathery forewings, exposing the abdomen. The four nymphal stages resemble the adults except that they are smaller and wingless. Earwigs are largely nocturnal and hide during the day, often in large aggregations, in cracks and crevices, or under bark. When disturbed they move quickly.

**Life Cycle and Damage**

Adults overwinter in the ground in earthen cells. Clutches of eggs laid in cells in the top 5 cm of soil in the fall and spring are opened by females in spring to release the young nymphs. Adults are semi-social and tend their young for a period of time after birth; nymphs later disperse to secure locations under rocks, in crevices, or under bark. Adults develop by mid-summer and there is only a single generation per year. Earwigs are nocturnal and feed at night on dam-
aged fruit, tender plant tissue, small soft bodied insects, and other foodstuffs. They rarely damage grape leaves.

**Monitoring and Spray Thresholds**

Rolled up newspaper or cardboard, or flat boards placed on the ground, can be used to monitor earwigs. It might be possible to use these same refuges to enhance their numbers in wine grapes.

**Biological Control**

Birds, large spiders, wasps and ground beetles prey on earwigs. *Earwigs, in turn, feed on the eggs and small nymphs of leafhoppers and on other soft bodied insects.*

**Cultural Control**

Tillage in spring and fall will destroy some of the earthen cells. For table grapes, mowing, tillage, removal of loose bark from vine trunks, and sanitation will remove hiding places and help reduce earwig numbers. They can be kept out of small plantings of table grapes by applying sticky tape or commercial Stickum® around trunks and posts. Washing table grapes after harvest will help dislodge many earwigs. They are less likely to seek refuge in table grape varieties with loose, open fruit bunches.

**Chemical Control**

Earwigs are sensitive to most insecticides and sprays applied for the control of other grape pests will usually also provide control of earwigs. Commercial earwig baits are also available.

**Root Weevils**

Black Vine Weevil, *Otiorhynchus sulcatus* F.

Raucus root weevil, *O. raucus* F.

**Life Cycle and Damage**

Black vine weevil and raucus root weevil spend the winter in the soil as pale, legless larvae feeding on the roots of a wide range of plants. It is not known if they feed on the roots of grapevines to any extent.

Adult black vine weevils feed during late spring and summer on grape clusters and leaves and sometimes girdle fruit clusters.

They are active only at night and remain hidden under plant debris or loose bark during the day, making detection difficult. Raucus root weevil adults, in contrast, can often be found feeding on the buds of grapes at twilight or on overcast days. Earlier emergence of adult raucus root weevil in spring that coincides with the development of grape buds accounts for the greater damage by this species as compared with the black vine weevil. Although localized and usually confined to the edges of vineyards, infestations of raucus root weevil have occurred occasionally in large numbers that required control. The raucus root weevil was reported from the Okanagan Valley for the first time in 1979 and there have been three localized outbreaks documented since that time.

**Identification**

The flightless adults of both species have long, broad ‘snouts’ typical of weevils (Coleoptera: Curculionidae). The thick, roughened front wings are fused together. Adults range in size from 5.5-7.5 mm for the raucus root weevil and 7.5-10 mm for the black vine weevil. In addition to their slightly smaller size, raucus root weevil can be differentiated from black vine weevil by their generally lighter, patchy appearance and often a distinct difference in colour between the head/thorax and the elytra that cover the abdomen (1f).

Black vine weevil adults are a uniform black colour. Unlike adult raucus root weevil that damage buds, leaves with deeply notched edges not associated with damage from other chewing pests are an indicator of black vine weevil feeding. Traps for both species can be made from cardboard loosely wrapped around vine trunks where the weevils will hide during the day. Adults can also be collected on beating trays placed beneath canes and cordons that are hit sharply with a stick or bat. Care needs to be taken, however, as adults often fall to the ground as they are approached.

**Click Beetle**

*Coleoptera: Elateridae*

Click beetles were formerly considered to be
important pests of grapes in British Columbia, but extensive study in Okanagan vineyards over a number of years has shown that most of the damage formerly attributed to click beetle adults was in fact caused by half grown cutworm larvae. Several species will feed occasionally on buds of grapes, but control is usually not warranted. Click beetles are also not considered pests of grape in Washington State.

**Identification**

Click beetle larvae, called wireworms, have long, slender, hard bodies that are a light tan to a light orange-brown in colour and appear somewhat shiny and polished. Their short legs and cylindrical slender bodies allow them to burrow through the soil to feed on the roots of plants. Adult beetles have elongate somewhat flattened bodies usually in shades of brown or black (1e). The segment behind the head has small projections that point backward on each side of the body. Adults of most species are around 1 to 1.5 cm in length but some are significantly larger.

When adults are placed upside down in the palm of a hand or gently held between finger and thumb they will rapidly flex their bodies at both ends and attempt to right themselves or escape with a characteristic clicking action.

**Life Cycle and Damage**

Adults that have spent the winter under debris and leaves emerge with the arrival of warm weather in spring to feed on the tender buds and leaves of a wide range of plants, including occasionally grapes. They are often quite common on flowers where they feed on soft tissues and pollen. After feeding for a short time, females begin to deposit eggs into the soil. In two or three weeks the eggs hatch into small slender ‘worms’, the larvae, that develop in the soil throughout the summer. Click beetles can persist for a long time without feeding and they often require more than one year to complete development to the adult stage.

Larvae are not known to damage the roots of grapes to any extent, preferring to feed on the roots of weeds or grasses. As for cutworm larvae, feeding of adults on buds of grape causes the buds to die. Yields can be reduced and ripening of fruit can be uneven and delayed. Little damage occurs after bud break.

**Monitoring and Spray Thresholds**

Adult click beetles feed on buds so infrequently that monitoring for these pests is generally not warranted. The presence of adult click beetles in a vineyard does not indicate feeding or correlate with damage. There are no established thresholds in British Columbia relating click beetle numbers to bud damage. To determine if bud damage is due to click beetles or cutworm larvae, it is best to inspect vines with the aid of a flashlight soon after dark on warm spring nights.

**Biological Control**

Little information is available on the biological control of click beetles in British Columbia vineyards.

**Cultural Control**

As for climbing cutworm, the presence of broadleaf weeds in spring helps reduce damage to grapes.

**Chemical Control**

Chemical control of click beetles is generally not warranted and none are registered specifically for adults on bearing vines. Sprays for cutworm larvae should help control adult click beetle.

**Miscellaneous Insect Pests**

In addition to the major and minor grape pests described in previous sections, a number of insects attack grapes only very rarely and are generally not of economic importance in British Columbia. Following are brief descriptions of a few of these occasional pests.
Wood-Boring Beetles

Two or three species of wood-boring beetles can occasionally infest older canes and vine trunks. Depending on the species, adults are brown to black, cylindrical in shape, and from 5-10 mm in length. They have hardened forewings typical of beetles and indistinct body sections. Adults burrow into the spurs and canes at the base of new shoots, causing them to wilt or break. The pale larvae or grubs burrow through living or dead tissues. Damage usually occurs on older or diseased vines.

Grasshoppers

Orthoptera: Acrididae

Adults and nymphs of several species of grasshopper feed on grape leaves, usually beginning in late summer when preferred hosts begin to dry up. Feeding by these well-known, long-legged jumping insects consists of chewing damage to leaf margins, most often on lower leaves. Grasshoppers suffer from pathogenic diseases when populations are high. Preying mantids, toads, and many species of birds feed on adults, while smaller nymphs are attacked by spiders and a number of predatory insects such as ground beetles.

Whitefly

Hemiptera: Aleyrodidae

Several species of whitefly occasionally infest grapes. Overwintering whitefly are relatively rare in British Columbia. Some sub-tropical and tropical species, such as the common greenhouse whitefly, Trialeurodes vaporariorum (Westwood), infest outdoor plants each season from infested greenhouses, house plants, and vegetable and ornamental transplants. Adults are small, around 2 mm in length, with two pairs of wings that are covered with a fine, white powder (3b). They somewhat resemble small white moths. The minute crawlers are mobile for only a few hours until they settle and develop into nearly translucent, scale-like immobile nymphs. The ‘scales’ have a fringe of waxy material projecting from the bottom edge (3g); the distribution and numbers of the projections can assist in species identification. The pupal stage that develops inside the case on the underside of leaves is somewhat bulbous and appears segmented. There are several generations per year. Whitefly feed by sucking juices from the leaves, much as scale insects do, and they also produce large amounts of sugary honeydew. Populations are controlled by parasitic Encarsia wasps that turn the scales black, by several predatory insects, and by pathogenic fungi.

Western Grape Rootworm

Bromius obscurus (L.)

The western grape rootworm is a widespread pest of grapevines in Europe and California, but damage to grapes in British Columbia was not reported until recently when an outbreak occurred in the Kelowna area. Fully grown larvae are C-shaped white grubs about 7 mm long with yellowish brown heads and dark mouthparts. Adult beetles are about 4 mm long and somewhat hairy with nearly black heads and dark bodies (5d insert). Adults, which are all females, start emerging in May and feed on the expanding leaves of grapes, causing long slit-like holes (5d). Larvae initially feed on root hairs before tunneling into the roots of grapevines. Feeding by western grape rootworm larvae is known to cause the death of vines in California due to direct damage to the roots and entry of secondary pathogens. Additional information is available at http://www.al.gov.bc.ca/cropprot/bromius_obscurus.pdf.

Minor Cicada

Platypedia minor Uhler

Adult minor cicada (Hemiptera: Cicadidae) are about 2 cm long with two pairs of large, transparent wings held roof-like over the body (3b). The pale, wingless nymphs have enlarged front legs that they use to dig through soil. Cicadas are mostly known from the loud buzzing or clicking noises that males produce on hot summer days. They are difficult to locate and will move around to the opposite side of trees or posts when approached. Damage to individual shoots occurs occasionally when females deposit eggs into canes with their long, saw-toothed ovipositor. Deposition of several eggs into a cane can weaken it or cause it to break. The oviposition damage is recognizable by the slivers of wood protruding from a series of punctures. Newly hatched nymphs drop to the
ground to spend two to three years feeding on the roots of plants. It is not known if they feed on the roots of grapes.

Three-Cornered Alfalfa Treehopper
*Spissistilus festinus* (Say)

Adult three-cornered alfalfa treehopper (*Homoptera: Membracidae*) are green in colour and about 6 mm in length. The pronotum is elongated to the sides and extends to the rear of the abdomen so that they look somewhat triangular when viewed from the front. Lacking the large side projections, the buffalo treehopper, *S. bisonia* Kopp & Yonke, which also reportedly feeds on grapevines, has the hump-backed appearance of the American Bison after which it is named. Mainly restricted to the south end of the Okanagan Valley, damage caused by the three-cornered alfalfa treehopper can somewhat resemble leafroll virus. In summer late instar nymphs will produce a series of feeding punctures around leaf petioles or stems (3a), resulting in decreased flow of water and nutrients and eventual reddening of leaves beyond the feeding site. The initial feeding punctures coalesce into a darkened band of sunken tissue encircling the stem or leaf petiole. Although damage is often restricted to lateral shoots and generally not of economic concern, growth of newly planted vines could be stunted in areas with lots of alfalfa or other suitable host plants.

### Prevention of New Grape Pests

Provincial Entomologists (currently Susanna Acheampong, Kelowna; Tracy Hueppelsheuser, Abbotsford), AAFC researchers, and Canadian Food Inspection Agency (CFIA) regulators and inspectors contribute to the prevention of the arrival of new invasive pests of grapevines into British Columbia. Importation of host plant material from areas known to harbour these pests is regulated and plants inspected upon arrival or at the source nursery. Information is provided to industry members and the general public through talks, identification factsheets and brochures. It is inevitable that new economically important pests of grapevines will arrive in our region from elsewhere. Growers can help prevent the establishment of new alien pests by knowing the species currently found in British Columbia and reporting new pests and unfamiliar damage. The list of potential grape pests that might arrive in British Columbia from elsewhere is very long. Following is a list of a few economically important ones and sources for additional information.

#### Brown Marmorated Stink Bug

*Halyomorpha halys* Stål

The brown marmorated stink bug (*Hemiptera: Pentatomidae*) was first found in eastern North America in 2001 and more recently in Washington and Oregon. It feeds on a wide range of vegetables, field crops, ornamental trees and shrubs, and fruit crops including grapes. Similar in size and shape to some of our native stink bugs, first instar nymphs are bright red marked with black, while adults are dark brown with white banding on the antennae and legs. Adults are a little more than 1.5 cm in length. In areas where they have become established they are often found feeding in close proximity to one another. Adults aggregate in large numbers in the fall to spend the winter in protected areas or invade houses. Damage results from feeding punctures and removal of fluids from the plant, resulting in shrunk, distorted buds, shoots and fruit.

For additional information on this pest see the brochure ‘Brown Marmorated Stinkbug’, BCMAL, posted at [http://www.agf.gov.bc.ca/cropprot/bmsb_alert.pdf](http://www.agf.gov.bc.ca/cropprot/bmsb_alert.pdf)

#### Light Brown Apple Moth

*Epiphyas postvittana* (Walker)

The light brown apple moth, which is native to the cooler, wetter coastal regions of Australia, was found in parts of California in 2007. As for other members of this moth family (*Tortricidae*), it has a wide host range that includes ornamentals, fruit trees, berries, and grapes. There likely would be two generations under our conditions. Larvae that grow to about 1 to 1.8 cm in length are pale to medium green with
a tan coloured head. Adult moths are medium tan often with oblique markings of darker tan to brown. Partly grown overwintering larvae feed on grape buds in spring and continue to feed on developing shoots, tender leaves and flower clusters as they mature. Damage to fruit later in the season reduces the crop and allows the entry of disease organisms.

For additional information on the light brown apple moth visit UC IPM Online, ‘http://www.ipm.ucdavis.edu/PMG/r2302303011’.

**European and American Grape Berry Moths**

The European grape berry or vine moth, *Lobesia botrana* (Denis & Schiffermüller), and the American grape berry moth, *Paralobesia (=Endopiza) viteana* Clemens, are similar in appearance and biology and cause similar damage to grapes. The former was first reported in Napa County, California, in 2009, while the latter from eastern North America is now found in parts of western Colorado. The slender larvae reach a maximum size of 1 to 1.5 cm and are green to light purplish with a darker contrasting head and first thoracic segment. The adult moths are generally less than ¾ of a centimeter long and of a medium brown colour mottled in patches of lighter or darker brown, black grey or blue depending on the species. Both are best identified by the damage caused by larvae feeding on flowers and fruit. Larvae of the 1st generation feed on developing buds, flowers and small berries and produce a significant amount of webbing. Larger 1st and 2nd generation larvae burrow into and feed on the fruit, leaving hollowed out skins and seeds. Grape bunches become contaminated with frass and webbing and are subject to fungal infections.


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**Additional information on grapevine pests and their control:**


UC Integrated Viticulture Online. University of California website. Main Menu>Viticultural Information>Insects and Mites.


Revised 2013. (Free downloadable PDF version at: cru.cahe.wsu.edu/CEPublications/eb0762/eb0762.pdf).
